

Dispersive liquid-liquid microextraction for the preconcentration and spectrophotometric determination of copper(II) in blood serum sample using sodium diethyldithiocarbamate as the complexing agent

Reza Emamali Sabzi¹, Naimeh Mohseni¹, Morteza Bahram^{1*}, Mahmoud Rezazadeh Bari²

¹ Department of Chemistry, Faculty of Science, Urmia University, Urmia, Iran

² Department of food science and technology, Faculty of agriculture, University of Urmia, Urmia, Iran

Abstract: A dispersive liquid-liquid microextraction (DLLME) process based on the complexation reaction of Cu(II) with diethyldithiocarbamate (DDTC) from aqueous solutions was investigated. The effect of various experimental parameters in the extraction such as, dispersive solvent, extracting solvent, the volume of extraction and disperser solvent, pH of the aqueous solution, ionic strength and extraction time were optimized using one variable at a time method, and the analytical characteristics of the method were obtained. Under the optimum conditions the calibration graph was linear over the range 0.01 to 0.1 $\mu\text{g mL}^{-1}$ of Cu(II) ion with a correlation coefficient of 0.994. The limit of detection ($S/N = 3$) was $8.6 \times 10^{-3} \mu\text{g mL}^{-1}$. Relative standard deviation (RSD) for 7 replicate determinations of 0.08 $\mu\text{g mL}^{-1}$ Cu(II) was 3.3%. In this work, the concentration factor of 20 and also the improvement factor of 33 were reached. The interference effect of some anions and cations was also tested. The extraction method has been successfully applied to the determination of copper in human blood serum sample.

Keywords: Copper; Sodium diethyldithiocarbamate; Dispersive liquid-liquid microextraction; Spectrophotometric determination; Blood serum sample.

Introduction

Copper is an essential micronutrient that has a crucial role in a variety of biological processes indispensable to sustain life¹. At the same time, higher concentrations of copper can be toxic and cause stomach and intestinal distress, liver and kidney damage, and anemia. The US Environmental Protection Agency (USEPA) and the World Health Organization (WHO) recommended maximum allowable concentration of 1.3 and 2 mg dm^{-3} , respectively for copper in drinking water^{2,3}. Since it has had numerous applications in the industry, copper pollution in the environment can occur that may cause toxic effects to living organisms in natural waters or humans. Therefore, circulating levels of this metal in the plasma must be kept under tight regulation.

The direct determination of heavy metals including copper at trace levels using various modern instrumental methods including flame atomic absorption spectroscopy (FAAS),

*Corresponding author: Morteza Bahram

E-mail address: m.bahram@urmia.ac.ir

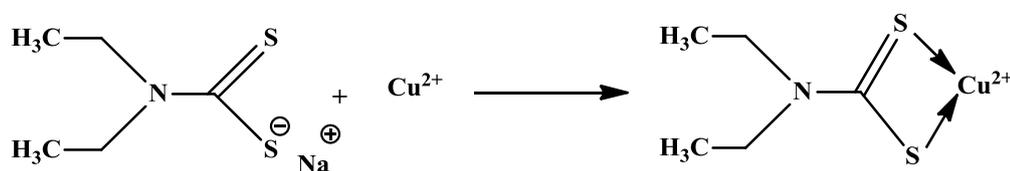
DOI: <http://dx.doi.org/10.13171/mjc.3.6.2015.01.10.14.50>

inductively coupled plasma-optical emission spectrometry (ICP-OES) and inductively coupled plasma-mass spectrometry (ICP-MS) has been unsuccessful due to particularly their low concentrations and matrix effects. However, preconcentration and matrix elimination is usually required for the determination of low concentrations of the metal ions in environmental samples⁴.

Various preconcentration and separation methods such as, liquid–liquid extraction (LLE)⁵⁻⁸, cloud point extraction (CPE)⁹⁻¹², solid phase extraction (SPE)¹³⁻¹⁶, ion exchange¹⁷ and co-precipitation^{18,19} hyphenated with different detection techniques, have been applied for extraction of trace levels of the heavy metal ions including copper(II) from different biological and environmental samples. However, some of these methods suffer from inconveniences such as, lengthy separation, limitation of the volume of sample solution investigated, time consuming, multi stage, yielding unsatisfactory preconcentration factors and the use of large quantities of organic solvents.

In 2006, Assadi and co-workers developed a novel liquid phase microextraction technique, named dispersive liquid-liquid microextraction (DLLME)^{20,21}. It is a simple and fast microextraction technique based on ternary component solvent systems. This method needs microliter volumes of extraction solvents and little amounts of a disperser solvent. The extraction solvent is an organic solvent such as chlorobenzene, chloroform or carbon disulfide with high density and the disperser solvent is an organic solvent, such as methanol, acetone or acetonitrile that is miscible with both extraction solvent and water. When the mixture of extraction phase and disperser is rapidly injected into the sample, a cloudy solution is formed. Since the surface area between the extracting solvent and the aqueous sample is very large, thus the equilibrium state is achieved quickly and the extraction is independent of time. In fact, this is the main advantage of DLLME. After centrifugation, the fine particles of extraction solvent sedimented in the bottom of the conical test tube. The sedimented phase is removed manually by a syringe and analyzed by various analytical techniques including, high-performance liquid chromatography (HPLC)²², electrothermal atomic absorption spectrometry (ET-AAS)²³⁻²⁵, FAAS²⁶⁻²⁹, ICP-OES³⁰, spectrophotometric analysis^{31,32} and energy-dispersive X-ray fluorescence spectrometry³³.

Diethyldithiocarbamate (DDTC) is a classical bidentate ligand that forms complexes with many metal ions including Cu^{2+} , Co^{2+} and Ni^{2+} . The complexing properties of this ligand is directly connected with the presence of two donor sulphur atoms (Scheme 1)³⁴. DDTC metal complexes are insoluble in water but soluble in organic solvents, therefore a solvent-extraction step is necessary.



Scheme 1. Chemical structure of sodium diethyldithiocarbamate in its free and complexed form.

X.Wen et al³⁵ have applied dispersive liquid–liquid microextraction combined with UV–vis spectrophotometry for determination of cadmium and copper in water and food samples. The limit of detection in this work for Cu(II) determination was reported as $0.5 \mu\text{g L}^{-1}$.

In this report, dispersive liquid-liquid microextraction method has employed for the preconcentration and spectrophotometric determination of Cu(II) using DDTC as the complexing agent. The most important aim of this work was clearly study of the related parametrs in dispersive extraction method. In the previous work³⁵ the calculated LODs for two studied cations were very ambiguous. Also in proposed method the applicability of DLLME for determination of Cu(II) in human plasma was studied. The experimental parameters were optimized using one variable at a time method. The proposed method is simple, efficient and rapid and it uses extracting solvent at a microliter level. Furthermore, a well-known and cheap ligand diethyldithiocarbamate was utilized. The extraction method has been successfully applied to the determination of Cu(II) in blood serum sample.

Experimental Section

Reagents

All experiments were performed with analytical reagent grade chemicals. All chemicals such as sodium diethyldithiocarbamate, methanol, ethanol, chloroform, dichloromethane, acetonitrile and tetrahydrofuran were obtained from Merck (Darmstadt, Germany) and used without any purification. Individual stock solution of Cu(II) at a concentration of $1000 \mu\text{g mL}^{-1}$ was prepared by dissolving appropriate amount of salt in distilled water. Standard solution of $1 \mu\text{g mL}^{-1}$ Cu(II) was prepared daily by a suitable dilution of stock solution with distilled water. Sodium diethyldithiocarbamate 0.05 M was prepared in distilled water. Acetate buffer (pH = 5) was used for pH adjustment.

Apparatus

UV-visible spectra were measured by using Shimadzu UV-2550 double-beam spectrophotometer with 1-cm quartz cells. Phase separation was assisted using Urum-Tadjihiz centrifuge (Urmia, Iran) centrifuge. A Metrohm model 713 pH-meter with a combined glass electrode was used for pH measurements.

General analytical procedure

Appropriate amount of Cu(II) solution was transferred into a 12 mL test tube with conical bottom, 1 mL acetate buffer and 1 mL DDTC solution were added. By using a 5-mL syringe, 1 mL methanol containing 200 μL chloroform was added to the above solution. Chloroform was dispersed in all parts of sample and no need to homogenize the sample. The mixture was immediately centrifuged for 5 min at 1000 rpm. The sedimented phase was quantitatively transferred to another test tube and allowed to evaporate at room temperature. Finally the residue was diluted with 0.5 mL ethanol and its absorbance was measured at 440 nm.

Analysis of blood serum samples

Blood sample was taken from a healthy volunteer, centrifuged at 3000 rpm for 10 min and serum fractions were collected. Precipitation of proteins was carried out by adding phosphoric acid (2 mol L^{-1}) to the serum sample. The pH of the samples was adjusted at 5. 1 mL methanol and 200 μL of tetrachloride carbon were added to the samples. The tube was centrifuged for 5 minutes and the clear supernatant liquid was used. Samples of the supernatant liquid (each of 2 mL) were fortified with various concentrations of Cu(II). Then the analysis was followed up as indicated in the general analytical procedure.

Results and Discussion

Figure 1 shows the absorption spectra for the individual metal complex in organic phase against reagent blank. The addition of an aqueous solution of DDTC to a slightly acidic solution of Cu(II) ion produces a brown colloidal suspension of Cu(II)-DDTC which is water insoluble but easily dissolved in several organic solvents.

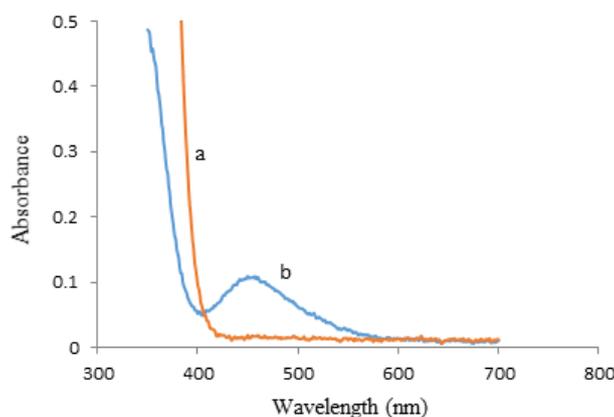


Figure 1. The absorption spectra of (a) diethyldithiocarbamate and (b) its complex with Cu(II) at pH 5 after DLLME.

Dispersive liquid–liquid microextraction procedure

Selection of dispersive and extracting solvents

In this work, four solvents: methanol, ethanol, tetrahydrofuran and acetonitrile were studied as a dispersive solvent. The selection of the extracting solvent is critical and should meet the following characteristics: (1) it should dissolve the analyte better than water, (2) it should be heavier than water and (3) it should form tiny droplets when it is added to the aqueous solution of analyte along with a dispersive solvent. Almost all of the suitable extracting solvents are chlorinated. In this work, chloroform, carbon tetrachloride and dichloromethane were investigated. For all combinations of dispersive and extracting solvents the absorbance of the Cu(II)-DDTC extracted to the organic phase was measured at 440 nm against the reagent blank.

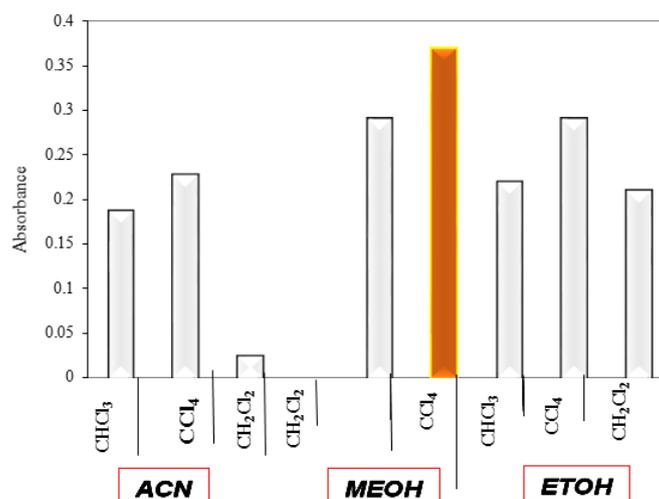


Figure 2. Selection of extracting and dispersive solvents in DLLME. Conditions: Sample, 5 mL Cu(II) $0.1 \mu\text{g mL}^{-1}$; volume of dispersive solvent, 1 mL; volume of extracting solvent, 200 μL ; buffer, 1 mL acetate buffer ($C = 1 \text{ M}$, $\text{pH} = 5$) and DDTC solution, 1 mL 0.05 M .

As it is shown in Figure 2, maximum extraction efficiency was obtained for the combination of carbon tetrachloride as extracting solvent with methanol as dispersive solvent. Therefore, the combination of carbon tetrachloride and methanol was selected for further studies.

Dispersive solvent volume

Methanol as a dispersive solvent in different volumes in the range 0–2.5 mL along with 200 μ L carbon tetrachloride as an extracting solvent was used for extraction of Cu(II)-DDTC complex using the DLLME procedure. The results (Figure 3) show that in the case of 1 mL methanol the highest absorbance signal was obtained. Therefore, methanol volume of 1 mL was selected as being optimal for all subsequent experiments.

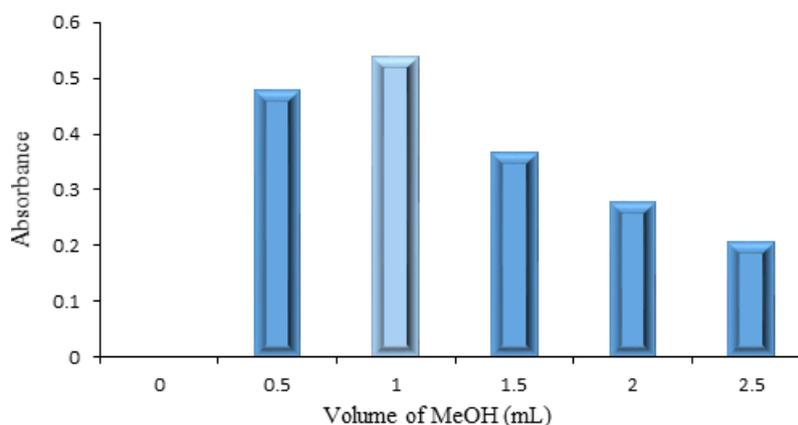


Figure 3. Dispersive solvent volume study. The absorbance of extracted phase vs. dispersive solvent volume. Conditions are the same as Figure 2.

Extracting solvent volume

Extracting solvent (carbon tetrachloride) volumes on the absorbance and recovery of analyte, in the range of 50–300 μ L was investigated. Since the absorbance signal was highest at 200 μ L of CCl_4 as extracting solvent (Figure 4), this was used in further experiments.

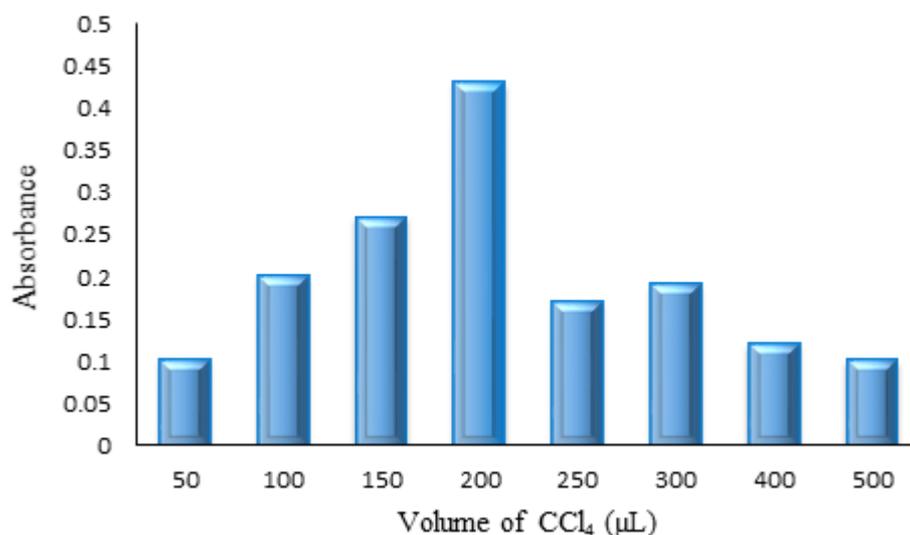


Figure 4. Selection of extracting solvent volume. The absorbance of extracted phase as a function of extracting solvent volume. Conditions are the same as Figure 3.

Effect of pH

One of the most important parameters for obtaining high recovery values was the reaction pH between the ligand and copper. DLLME was performed at different pH values in the range of 1–11. Figure 5 shows that the absorbance signals increase with an increase in the solution pH up to a maximum value of 5 and then decreases with further increases in the solution pH. At very alkaline pH values, the Cu(II) ions undergo hydrolysis and precipitate in the hydroxide form and measurements at these pH values mask any result. At acid pH the decrease in absorbance can be attributed to the protonation of the complexing agent. Thus, there is competition between the protons and the Cu(II) ions at more acid pH values. Therefore, pH 5 was selected for all subsequent experiments.

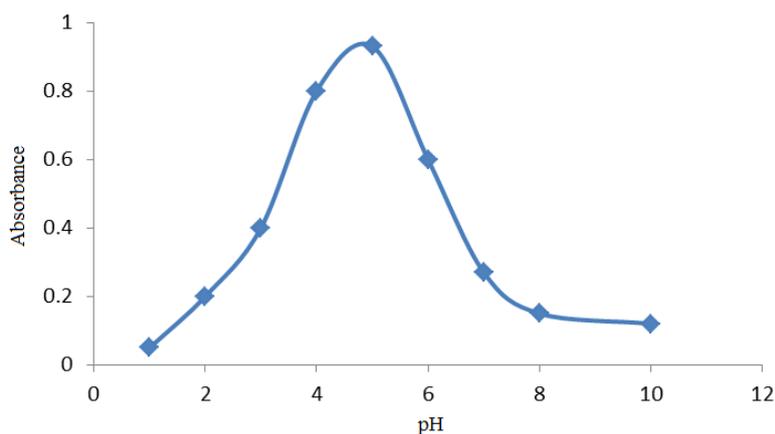


Figure 5. The effect of pH on the absorbance of Cu-DEDTC complex in extracted phase.

Sample volume

Different volumes of analyte solution (5–20 mL) were used to study the effect of sample size with a constant volume of extracting solvent (200 μ L) and dispersive solvent (1 mL). As it can be seen in Figure 6, with a large sample size (20 mL) no sedimented organic phase was obtained. By increasing the sample volume from 5 to 15 mL, the volume of sedimented phase and recovery decreased. Therefore, sample volume of 5 mL was selected for the following studies.

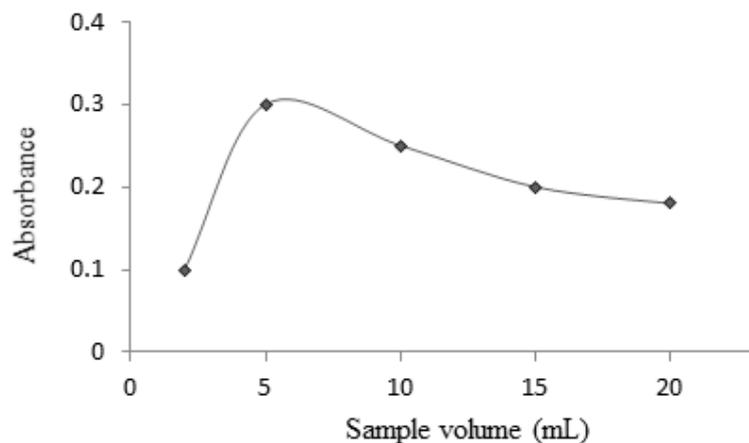


Figure 6. The effect of incubation time prior to centrifuge on the complex formation.

Study of other parameters

Other parameters such as DDTC concentration, salting out effect and reaction time were investigated. To determine the suitable ligand concentration, different concentrations of ligand in the range of 10^{-4} - 10^{-2} mol L⁻¹ was added to 5 mL of the analyte solution containing 0.1 mg L⁻¹ Cu(II) at pH 5. These samples were analyzed according to the procedure. As it is shown in Figure 7, absorbance signals increased up to 10^{-3} mol L⁻¹ and then increasing ligand concentration did not influence the extraction recovery. Therefore, a ligand concentration of 10^{-3} mol L⁻¹ was selected as being optimal for all subsequent experiments.

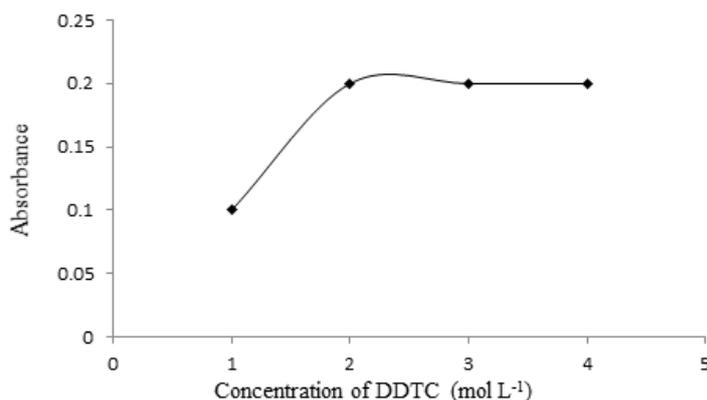


Figure 7. The effect of sample volume on the extraction efficiency at optimum conditions.

Also, the influence of ionic strength on the extraction was studied with sodium chloride as a salting out agent at the range of 0.1-1 mL of saturated NaCl solution. The highest absorbance signal was obtained at 0.3 mL of saturated NaCl solution (Figure 8). It was found that by increasing sodium chloride, the volume of sedimented phase decreases.

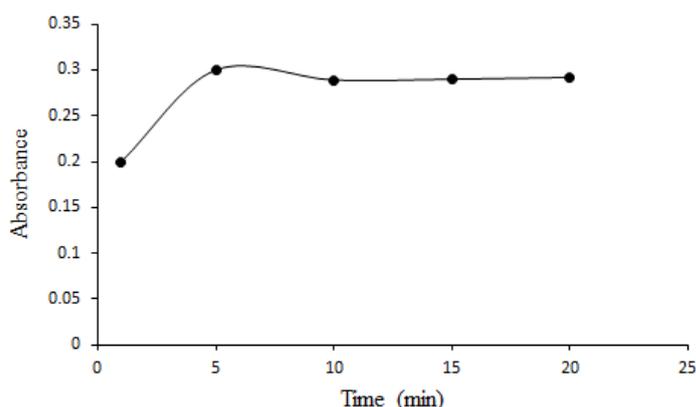


Figure 8. The effect of complexing agent diethyldithiocarbamate on the absorbance of the extracted phase.

Extraction time also was studied over the range 1–20 min (Figure 9). It was defined as the time spent between the addition of extraction solvent (carbon tetrachloride) dissolved in dispersive solvent (methanol) and centrifuging. Since the surface area between the extracting solvent and the aqueous sample is very large, thus the equilibrium state is achieved quickly and time did not affect on the recovery and sediment phase volume.

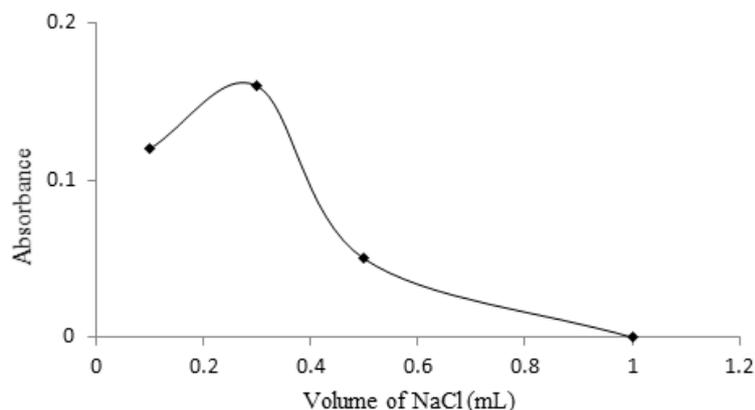


Figure 9. The effect of saturated NaCl solution on the extraction of Cu(II)-DDTC complex.

Analytical Characteristics

Table 1 summarizes the analytical characteristics of the optimized method, including regression equation, linear range, limit of detection, reproducibility, preconcentration, and improvement factors. Under the optimum conditions the calibration graph was linear over the range 0.01 to 0.1 $\mu\text{g mL}^{-1}$ of Cu(II) ion with a correlation coefficient of 0.994. The limit of detection, defined as $C_L = 3 S_B/m$ (where S_B and m are standard deviation of the blank and slope of the calibration graph, respectively) was $8.6 \times 10^{-3} \mu\text{g mL}^{-1}$. Because the amount of Cu(II) ions in 10 mL of initial sample solution is measured after DLLME in a final volume of 0.5 mL, the solution is concentrated by a factor of 20. The improvement factor, defined as the ratio of the slope of the calibration graph for DLLME method to the slope of the calibration graph without preconcentration was 33. Relative standard deviation (RSD) for 7 replicate determinations of $0.08 \mu\text{g mL}^{-1}$ Cu(II) was 3.3%.

Table 1. Analytical features of DLLME of Cu(II) ions.

Regression equation ^a	Abs = 1.982C + 0.003
Regression equation before extraction	Abs = 0.061C + 0.003
R ^{2b}	0.994
Linear range ($\mu\text{g mL}^{-1}$)	0.01-0.1
LOD ($\mu\text{g mL}^{-1}$) ^c	8.6×10^{-3}
Repeatability (RSD) ^d	3.3
Concentration factor	20
Improvement factor ^e	33

^a Concentration of analyte in $\mu\text{g mL}^{-1}$.

^b Squared regression coefficient.

^c Limit of detection for S/N = 3.

^d Relative standard deviation for 7 replicate determination of $0.08 \mu\text{g mL}^{-1}$ Cu(II) ions.

^e The ratio of the slope of the calibration graph for the microextraction method to that of the slope of the calibration graph without preconcentration.

Interference study

In order to study the selectivity of the proposed method, the effect of various cations and anions on the DLLME of Cu(II) ions was studied. An ion was considered as interferent, when it caused a variation in the absorbance of the sample greater than $\pm 5\%$ in comparison with the case in which interfering ion is absent. The results are summarized in Table 2. The results indicate that most of the cations and anions did not interfere even when present 1000-fold excess over analyte. As can be seen most ions studied do not have interfering effect at 1:100 ratio. Only Fe(III) interfered seriously, which was removed by addition of 5% triethanolamine (TEA) solution to the sample.

Table 2. Study of interfering ions.

Interfering ions	Tolerable concentration (analyte:interfering ion) ^a
Fe ³⁺	1:10
Co ²⁺ , Ni ²⁺ , Zn ²⁺ , Hg ²⁺ , NO ₃ ⁻	1:100
Cr ³⁺ , K ⁺ , NO ₂ ⁻ , EDTA, sodium citrate	1:1000

Concentration of analyte is $0.08 \mu\text{g mL}^{-1}$.

^a At this ratio no interfering effect was observed.

Application of the proposed DLLME method for determination of Cu(II) in blood serum

The extraction of the Cu(II) ions was conducted according to the procedure. For removal matrix effect in the serum sample, standard addition method was applied. The concentration of Cu(II) in blood serum was determined from intercept of the standard addition plots. The result is shown in Figure 10. The concentration of $4.5 \times 10^{-2} \mu\text{g mL}^{-1}$ was obtained for Cu(II) in the serum sample 1. Two other human plasma samples were analyzed and the results were presented in Table 3. The results indicate that the method can be successfully applied to recover Cu(II) ions in serum samples.

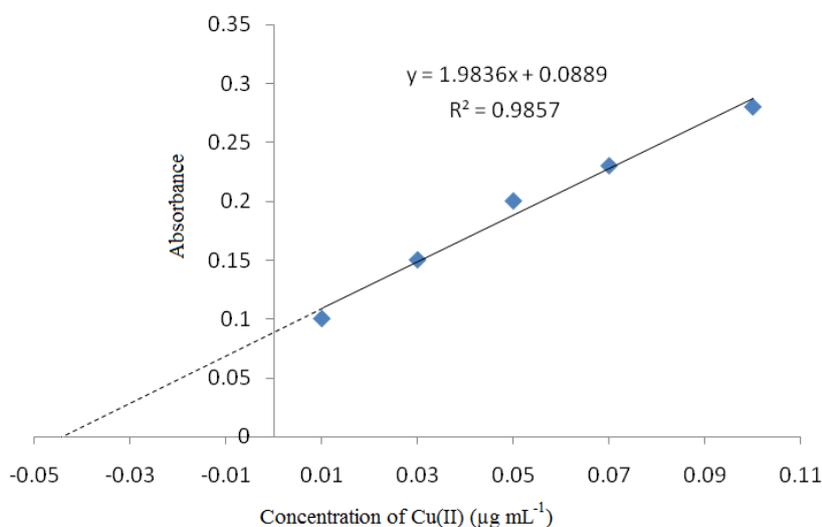


Figure 10. Standard addition plot of Cu(II) in human blood serum sample.

Table 3. Determination of Cu(II) in the plasma samples by the proposed method.

Samples	Added (ng mL ⁻¹)	Found (ng mL ⁻¹)	Recovery (%)
Sample 1	0	45	-
	40	97	105
Sample 2	-	<LOD	-
	40	41.2	103
	60	59	98.3
Sample 3	-	<LOD	-
	40	39.4	98.5
	60	57	94.5

Conclusion

A dispersive liquid-liquid microextraction for the preconcentration and spectrophotometric determination of copper(II) was used. The proposed method was based on the complexation reaction of Cu(II) with DDTTC from aqueous phase into the fine droplets of extraction solvent. Concentration factor and improvement factor for the target analyte were obtained about 20 and 33, respectively. The proposed extraction method was used for the quantitation of Cu(II) ions in blood serum sample. The method is simple, efficient and very rapid and it uses extracting solvent at a μL level.

References

- 1- D. L. de Romãna, M. Olivares, R. Uauy, M. Araya, *J Trace Elem Med Biol*, **2011**, 25, 3-13.
- 2- WHO, Copper in drinking-water: Background document for development of WHO guidelines for drinking-water quality. Geneva: World Health Organization (WHO/SDE/WSH/03.04/88), **2004**.
- 3- USEPA, Lead and copper monitoring and reporting guidance for public water systems. EPA-816-R-02-009. Washington, DC: Ground Water and Drinking Water Division, Water Programs, US Environmental Protection Agency, **2002**.
- 4- N. J. K. Simpson, *Solid Phase Extraction: Principles, Techniques and Application*, Marcel Dekker, USA, **2000**.
- 5- K. Wieszczycka, M. Kaczerewska, M. Krupa, A. Parus, A. Olszanowski, *Sep. Purif. Technol.*, **2012**, 95, 157-164.
- 6- S. H. Chang, T. T. Teng, N. Ismail, *Water Air Soil Pollut.*, **2011**, 217, 567-576.
- 7- D. Kara, M. Alkan, *Microchem. J.*, **2002**, 71, 29-39.
- 8- M. A. Farajzadeh, M. Bahram, S. Zorita, B. Ghorbani Mehr, *J. Hazard. Mater.*, **2009**, 161, 1535-1543.
- 9- P. Liang, J. Yang, *J. Food Compos. Anal.*, **2010**, 23, 95-99.
- 10- M. Bahram, S. Khezri, *Anal. Methods*, **2012**, 4, 384-393.
- 11- A. B. Tabrizi, *J. Hazard. Mater.*, **2007**, 139, 260-264.
- 12- E. L. Silva, P. S. Roldanb, M. F. Giné, *J. Hazard. Mater.*, **2009**, 171, 1133-1138.
- 13- E. M. Soliman, H. M. Marwani, H. M. Albishri, *Environ. Monit. Assess*, **2013**, 185, 10269-10280.
- 14- S. Chen, C. Liu, M. Yang, D. Lu, L. Zhu, Z. Wang, *J. Hazard. Mater.*, **2009**, 170, 247-51.

- 15- N. Farnad, K. Farhadi, N. H. Voelcker, *Water Air Soil Pollut.*, **2012**, 223, 3535-3544.
- 16- Q.M. Li, X.H. Zhao, K. Jiang, G.G. Liu, *Chinese Sci. Bull.*, **2007**, 52, 65-70.
- 17- A. Smara, R. Delimi, C. Poinsignon, J. Sandeaux, *Sep. Purif. Technol.*, **2005**, 44, 271-277.
- 18- H. Chen, J. Jin, Y. Wang, *Anal. Chim. Acta*, **1997**, 353, 181-188.
- 19- I. Komjarova, R. Blusa, *Anal. Chim. Acta*, **2006**, 576, 221-228.
- 20- M. Rezaee, Y. Assadi, M.R. Milani Hosseini, E. Aghaee, F. Ahmadi, S. Berijani, *J. Chromatogr. A*, **2006**, 1116, 1-9.
- 21- M. Rezaee, Y. Yamini, M. Faraji, *J. Chromatogr. A*, **2010**, 1217, 2342-2357.
- 22- A. Asghari, M. Ghazaghi, M. Rajabi, M. Behzad, M. Ghaedi, *J. Serb. Chem. Soc.*, **2014**, 79, 63-76.
- 23- R. E. Rivas, I. Lopez-Garcia, M. Hernandez-Cordoba, *Microchim. Acta*, **2009**, 166, 355-361.
- 24- E. Stanisz, A. Zgoła-Grześkowiak, *Talanta*, **2013**, 115, 178-183.
- 25- M. Amirkavei, S. Dadfarnia, A. M. Haji Shabani, *Quim. Nova*, **2013**, 36, 63-68.
- 26- D. Kantürer Acar, D. Kara, *Water Air Soil Pollut.*, **2014**, 225, 1864-1872.
- 27- M. A. Farajzadeh, M. Bahram, B. Ghorbani Mehr, J. A. Jonsson, *Talanta*, **2008**, 75, 832-840.
- 28- A. N. Anthemidis, K. -I.G. Ioannou, *Talanta*, **2009**, 79, 86-91.
- 29- H. Karimi, M. Ghaedi, A. Shokrollahi, H. R. Rajabi, M. Soyak, B. Karami, *J. Hazard. Mater.*, **2008**, 151, 26-32.
- 30- Y. Yamini, M. Rezaee, A. Khanchi, M. Faraji, A. Saleh, *J. Chromatogr. A*, **2010**, 1217, 2358-2364.
- 31- B. Horstkotte, M. Alexovič, F. Maya, C. M. Duarte, V. Andrich, V. Cerdá, *Talanta*, **2012**, 99, 349-356.
- 32- M. N. Uddin, Md. A. Salam, M. A. Hossain, *Chemosphere*, **2013**, 90, 366-373.
- 33- K. Kocot, B. Zawisza, R. Sitko, *Spectrochim. Acta Part B*, **2012**, 73, 79-83.
- 34- A. Hulanicki, *Talanta*, **1967**, 14, 1371-1392.
- 35- X. Wen, Q. Yang, Z. Yan, Q. Deng, *Microchem. J.*, **2011**, 97, 249-254.